

WEST Search History

DATE: Friday, April 02, 2004

Hide?	Set Name	Query	Hit Count
	<i>DB=PGPB,USPT,USOC,EPAB,DWPI; PLUR=YES; OP=ADJ</i>		
<input type="checkbox"/>	L1	6544741.pn.	2
<input type="checkbox"/>	L2	liang-Z\$.in. and randomized core	0
<input type="checkbox"/>	L3	liang-Z\$.in. and randomized	5
<input type="checkbox"/>	L4	0465219.pn. or 6458539.pn.	4
<input type="checkbox"/>	L5	(blocker near primer) or (blocking near primer)	140
<input type="checkbox"/>	L6	L5 and (oligonucleotide nea\$3 librar&)	0
<input type="checkbox"/>	L7	L5 and (oligonucleotide same librar&)	0
<input type="checkbox"/>	L8	L5 and (librar&)	0
<input type="checkbox"/>	L9	l5 and random\$	68
<input type="checkbox"/>	L10	64423493.pn.	0
<input type="checkbox"/>	L11	66423493.pn.	0
<input type="checkbox"/>	L12	6423493.pn.	2
<input type="checkbox"/>	L13	779290	13
<input type="checkbox"/>	L14	studier-W\$.in.	0
<input type="checkbox"/>	L15	Studier-W\$	0

END OF SEARCH HISTORY

End of Result Set☐ Generate Collection

Mar 20, 1996

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Basic Abstract Text (3) :

ADVANTAGE - Blocking primers prevent misannealing of specific primers to template sequences with which they are not exactly complementary. They also prevent annealing of oligonucleotide sequences produced in template repair processes from themselves acting as primers. A ddNTP at the 3'-end of the random or specific blocking primers prevents extension reactions. Enhancing reactions are performed using two primers for each nucleic acid strand. The primer closest to the 5'-end of the TNA may be extended whereas the primer closest to the 3'-end of the TNA (specific enhancing primer) has a ddNTP at its 3'-end and thus cannot act as an initiator of nucleic acid extension. The method produces extension prods. of known size.

L14 ANSWER 2 OF 6 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN

ACCESSION NUMBER: 1999-00412 BIOTECHDS

TITLE: Identification of biologically active DNA sequences;
by oligonucleotide expression in host cell; application in
e.g. Escherichia coli new tetracycline-resistance gene
promoter, beta-lactamase gene active site identification

AUTHOR: Horwitz M S; Loeb L A

PATENT ASSIGNEE: Univ.Washington

LOCATION: Seattle, WA, USA.

PATENT INFO: US 5824469 20 Oct 1998

APPLICATION INFO: US 1994-316415 30 Sep 1994

PRIORITY INFO: US 1994-361415 30 Sep 1994

DOCUMENT TYPE: Patent

LANGUAGE: English

OTHER SOURCE: WPI: 1998-582545 [49]

AN 1999-00412 BIOTECHDS

AB A method of obtaining an oligonucleotide that confers a predetermined biological function, regulation of expression or a biological activity of a protein, on a cell is new. This occurs by: cloning a heterogenous pool of oligonucleotides into an expression vector, where the cloned oligonucleotides are transcribed or act as regulatory sequences; introducing a random sample of the cloned oligonucleotides into a population of cells that do not exhibit the predetermined function; biologically selecting a subpopulation of cells exhibiting the predetermined function and then isolating the **central sequence** conferring this function. Also claimed are methods where the heterogenous pool of oligonucleotides contains oligonucleotides of specified length (54 bp or less) and composition. The process is used for e.g. identifying new forms of the Escherichia coli tetracycline-resistance gene promoter and the active site of the beta-lactamase (EC-3.5.2.6) gene. (24pp)

Generation of amplifiable genome-specific oligonucleotide
probes and **libraries**.

AUTHOR: **Brukner Ivan; Tremblay Guy A;**
Paquin Bruno

CORPORATE SOURCE: Universite de Montreal, Quebec, Canada..
ibrukner@hotmail.com

SOURCE: BioTechniques, (2002 Oct) 33 (4) 874-6, 878, 880 passim.
Journal code: 8306785. ISSN: 0736-6205.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 200303

ENTRY DATE: Entered STN: 20021026

Last Updated on STN: 20030327

Entered Medline: 20030326

AB Here we describe a process for the generation of **oligonucleotide libraries** representative of a given nucleic acid. Starting from at random pool of DNA oligonucleotides, the technique selects only those that hybridize to the nucleic acid template. This selection yields a highly specific **library** that represents an oligonucleotide image of the chosen template. The novel quality of this approach is the generation of amplifiable oligonucleotide probes that are of unique length and are easily subjected to differential selection. Here we apply this technique to produce different genomic **oligonucleotide libraries** and show that these genomic **oligonucleotide libraries** do not cross-hybridize. Differential selection of these genomic **oligonucleotide libraries** produces oligonucleotides that can be used in the identification, characterization, and isolation of nucleic acids.